

Effect of *Glomus mosseae* on growth of selected plant species

M Sandeepa G

Department of Biotechnology, S K University, Ananthapur, India

Received for publication: April 15th 2013; Revised: May 15th 2013; Accepted: May 20th 2013

Abstract: Effect of inoculating *Cajanus cajan*, *Arachis hypogaea*, *Ocimum tenuiflorum* and *Phyllanthus amarus* with mycorrhizal fungi (*Glomus mosseae*) was studied using potted seedlings. Vegetative growth and root length were compared between inoculated and non-inoculated plants. Result showed that there was a high mycorrhizal dependency of the selected plants on *G. mosseae*. Mycorrhizal fungal inoculated selected plants seedlings grew significant better compared to non-mycorrhizal controlled seedlings, and had increased height. *C. cajan*, *A. hypogaea* and *O. tenuiflorum* showed well established root colonization 87.6%, 86% and 93.5% respectively. Plant architecture and reproductive traits were changed by mycorrhizal fungi inoculation. Plants inoculated with *G. mosseae* invested more in the root development for effective nutrient absorption, which was the fundamental to achieve the maximum growth of the plants. The symbiosis of mycorrhizal fungi and rice would be of benefit to crop growth and grain yield.

Keywords: *Cajanus cajan*, *Arachis hypogaea*, *Ocimum tenuiflorum* and *Phyllanthus amarus*, *Glomus mosseae*. mvcorrhizal fungi. symbiosis.

Introduction

Mycorrhizal fungi are relevant members of the rhizosphere populations. Upon root penetration and colonization mycorrhizal fungi develop an external mycelium which connects the root with surrounding soil (Toro et al., 1997). Seven different types of Mycorrhiza were found, among them *Arbuscular Mycorrhiza* (Vesicular and Arbuscular Mycorrhiza) are the most common in occurrence (Mosse et al., 1983). These are found as key components of soil microbiota (Barea et al., 2004). AM fungi are obligately biotrophic organisms that live symbiotically with the roots of most plants. The establishment of a function symbiosis between AM fungi and host plants involves a sequence of recognition events leading to the morphological and physiological interaction of two symbions (Giovanetti & Sbrana, 1998). One of the most dramatic effects of these fungi on the host plant is the increasing phosphorus (P) uptake (koide, 1991) mainly due the capacity of the mycorrhizal fungi to absorb phosphate from soil and transfer it to the plant, (Asimi et al., 1980). Arbuscules are structures produced within the host plant cells by the AM fungi. These structures are responsible for the transfer of absorbed nutrients from the fungus to the plant. The present paper reports the influence of AM fungi on five different selected species of plants on plant height, mychorizal colonization root and shoot length, number and size of leafs and number of flowers.

Materials and Methods

To assess the effect of *Glomus mosseae* on the growth of different plant species, a pot culture experiment was done and plants were growing in earthen pots (30 cm diameter and 40 cm height) containing 7 kg of soil. The plants used are *Cajanus*

cajan, *Arachis hypogaea*, *Ocimum tenuiflorum* and *Phyllanthus amarus*.

Preparation of cultures: Mycorrhizal cultures were raised in the soil mixed with sand in the ratio of 1:1 and sterilized twice on alternate days. The cultures were raised employing the spores or sporocorps. For the raising of culture a technique provided by Gour and Adholeya, (1994) was followed. The top soil was removed upto the length of 3-5cm. AM inoculum *Glomus mosseae* (Nicolson and Gerdemann) was uniformly spread over the soil in a layer next soil was covered and seeds or seedlings are inoculated in soil above the inoculum. The *Glomus mosseae* culture was obtained from oil palm research centre, Peddavegi, Near Eluru, Andhra Pradesh, India.

Five seeds or seedling were sowed per pot. Pots were watered regularly. Observations per plant height, root and number and size of leaves, percentage of mycorrhizal colonization in roots and number of spores were recorded. The mycorrhizal colonization was determined using the method described by Philip and Hayman (1970). Mycorrhizal spores in the soil were estimated using wet sieving and decanting technique (Gerdemann and Nicolson, 1963). Plant height, root length, number of leaves and flowers, mycorrhizal colonization and number of spores present in soil were determined after 90 days after sowing.

Results and Discussions

Inoculation with fungi resulted in significant increase in plant height in *C. cajan*, *A. hypogaea* and *O. tenuiflorum* than control plants. These plants showed the significant increase in plant height 41.3

*Corresponding Author:

M Sandeepa,

Department of Biotechnology,
S K University, Ananthapur, India

cm, 23.03 cm, 31.0 cm than control plants 41.3 cm, 34.23cm, 30.6 cm respectively when inoculated with AM fungi. AM fungi also showed significant effect on these three plants root length and number of leaves than plants, the present findings has similarity with observations Dwivedi (2004) who inoculated AM fungi in wheat which showed maximum increase in plant height. *C. cajan*, *A. hypogaea*, *O. tenuiflorum* showed well established root colonization 87.6%, 86% and 93.5% respectively. Champawat (1992) reported that root colonization of mycorrhiza was 76% to 86% in chick pea plants grown in sterile soils. *Glomus mosseae* in the rhizosphere soils of wheat plant not only increased mycorrhizal root colonization (73%) but also helped in the nutrient uptake result in better growth of the plant and higher grain yield (Dwivedi, 2004). *A. hypogaea* culture plant showed significant

increase in mean number of flowers (38) than mean number of control plant (19.8). These finding showed that *Glomus mosseae* not only showed its effect on plants height, root length but also showed its effect on number and size of flowers and leaves also.

AM fungal inoculation in *P. amarus* did not show any significant effect on growth plant when compared to control. In some field studies mycorrhizal colonization has been shown to increase plant growth and survival, but these are many accounts of null or even negative effect (Gonigle and Fitter, 1998). The present findings in *P. amarus* showed less positive effect of AM inoculation. It may also be due to fewer interactions between fungus and plants at the cell wall and or middle lamellae level (Tester et al., 1987).

Plant name	Type	Shoot height (cm)	Root length (cm)	Number of leaves	Number of flowers	% of colonization	Number of spores
Cajanus cajan	Control	41.3	13	33
	Culture	79	16	74	...	87.6	29
Arachis hypogaea	Control	23.03	9	72	29.8
	Culture	34	11	109	38	86.0	10
Ocimum tenuiflorum	Control	31	14	51	4
	Culture	71.9	18	80	2	93.5	37
Phyllanthus amarus	Control	29	11	70
	Culture	29	10	84	42

References

- Asmi, S. Gianinazzi Pearson, V. and Gianinazzi, S. Influence of increasing soil phosphorus levels on interactions between vesicular arbuscular mycorrhiza and rhizobium in soya beans. *Canadian journal of Botany*, 1980, 58 vol, 2200-2205..
- Barea Jose-Miguel, Rosario Azcon and Concepcion Azcon-Aguilar. Mycorrhizosphere interactions to improve plant fitness and soil quality. *Antonie van Leeuwenhoek* 2004, 343-351..
- Champawat, R. S. Effect of different VAM fungi under varying levels of phosphorus on growth and nutrition uptake of chick pea. In: Jalali and Harichand (Eds.) *Trends in Mycorrhizal research*. 1992, Vol no. 160-161.
- Dwivedi O. P. Effect of mycorrhiza on winter wheat genotype C-306. *Mycopl pathol* 2004, 34, No 2.
- Gerdemann J. W. and Nicolson, T. H. Spores of mycorrhizal endogone species extracted from soil by wet sieving and decanting transe. *Br. Mycol Soc.* 1963, 46, 235-244.
- Giovanneppi and C. Sbrana. Meeting a non-host: the behaviour of AM fungi. *Mycorrhiza* 1998, 123-130.
- Gonigle, Mc, T. P and Fitter A. H. Growth and phosphorus inflows of trifolium repens L. with a range of indigenous vesicular arbuscular fungi. *New phytol* 1988, 08: 59-66.
- Gour. A and A. Adholeya. Mycorrhiza News. 1994, 6:10-11.
- Koide, R. T. Nutrient supply, Nutrient demand and plant response to mycorrhizal infection. *New phytologist* 1991, 117 vol. 365-386.
- Mosse, B., D. P. Stribley and F. Le Tacon. The ecology of mycorrhizae and mycorrhizal fungi. *Adv. Microbe Ecol.* 1983, 5:137-210.
- Phillip J. M and Hayman D. S. Improved procedures for clearing roots and staining parasitic and vesicular and arbuscular fungi for rapid assessment of infection. *Transactions of the British Mycological society.* 1970, 55:158-161.
- Tester. M, Smith S. E, Smith F. A. The phenomenon of non "mycorrhizal plants". *Can. J. Bot.* 1987, 65:419-431.
- Toro, M, Azcon, R. and Barea, J. Improvement of arbuscular mycorrhizal development by inoculation of soil with phosphate solubilizing rhizobacteria to improve rock phosphate bio availability (32 P) and Nutrient cycling. *Applied and Environmental Microbiology.* 1997, Nov, 4408-4412.

Source of support: Nil

Conflict of interest: None Declared